Adsorption of PCB's and DDT's on Membrane Filters— A New Analysis Method

David A. Kurtz Pesticide Research Laboratory Pennsylvania State University University Park, Pa. 16802

Numerous methods for the analysis of trace amounts of PCB mixtures and DDT isomers in water have been used and reported. EPA methods utilize pure hexane (BERCK 1953, KAHN and WAYMAN 1964, and EPA 1971). Several adsorption systems have been used to extract such compounds from water. They include activated carbon (ROSEN et al. 1959 and BREIDENBACH et al. 1966), urethane foam plugs (GESSER et al. 1971, UTHE et al. 1972, and UTHE et al. 1974) and Carbowax 4000 monostearate and undecane-coated Chromosorb W (AHLING et al. 1970). These methods have adsorption efficiencies from 80-100%. The foam plug method has given excellent recoveries from distilled water. However, when knowns were added to natural lake waters containing fine silt, lower recoveries were obtained (BEDFORD 1974). The PCB's were found to be adsorbed to the silt which passed through the plugs. This finding led to the work of this paper.

Adsorption methods have also been used in analyzing air samples for these compounds. Florisil packing has been used to collect PCB's and phthalates (GIAM et al. 1975), and urethane plugs also have been used for $\overline{\text{PCB}}$'s (BIDLEMAN et al. 1974). The urethane plugs in this latter study separated tri-, tetra-, and pentachlorobiphenyls from air at a 96-99% efficiency as compared to a 1-4% efficiency for glass fiber-A plugs.

The need still exists to analyze large quantities of water for PCB's and DDT's in ocean seas (CRAMER 1973, JENSEN et al. 1969, RISEBROUGH et al. 1968, and WOODWELL et al. 1971), state waters (EPA 1975a), and private ponds. Methods of increased sensitivity with convenient field sampling will be required (EPA 1975a).

This paper describes a new adsorption method by which PCB mixtures and DDT isomers and other chlorinated hydrocarbon residues can be efficiently separated from water sources. The adsorption medium is a membrane filter made from cellulose triacetate material. Field methods with simple apparatus can be used. The membrane system can be used both for analytical

purposes and for purifying contaminated sources. There is potential for its use in analyzing air samples.

MATERIALS AND METHODS

Metricel filter circles (Gelman, GA-6) with a pore size of $0.45~\mu$ and diameter of 47 mm were used without further treatment to separate PCB's and DDT's from These circles contained a small amount of starch cellulosic material used as a wetting agent. The flow of water through these was about 200 ml/min via water aspirator pumping. The filter holders were either stainless parabella (Gelman #4230, 1 liter capacity) or glass cone (VWR Scientific #28153020, 0.5 liter capacity). Hexane, acetone, cyclopentane, and acetonitrile (Burdick and Jackson distilled-in-glass) were used as received. Sodium sulfate (MCB, anhydrous, gran. reagent, lot 59) was heated at 400°C. Glassware was cleaned with a 5% solution of Contrad 70 cleaning agent (Scientific Products) heated to 70°C. Measuring pipets for dispensing standards were of the disposable type (Drummond microcap tubes). Column cleanup utilized 60/ 100 PR florisil (The Floridin Co.), activated at 1250°F and stored at 130°C.

Gas chromatographic analyses were accomplished with a Microtek MT-220 GLC fitted with a 5.5 foot $\frac{1}{3}$ inch o.d. glass "U" column packed with 1.5% SP-2250/1.95% SP-2401 (similar to 0V-17/QF-1) on 100/120 Supelcoport packing (Supelco, Inc.), operated at 215°C oven temperature, 240°C inlet and nitrogen flow of 60 ml/min. The 63 Ni electron capture detector was held at 330°C and had a 20 ml/min nitrogen purge. The column gave from 3000 to 6000 theoretical plates efficiency for p,p'-DDT. The electrometer sensitivity was 1.6 x 10^{-9} amps full recorder scale background current. Analyses were quantified with an Infotronics Model CRS-100 area integrator.

Standards were prepared by adding μl quantities (10-100) of Aroclor (PCB) mixtures and DDT isomers dissolved in isooctane directly on 3.00 liters of double-distilled water contained in one-gallon Burdick and Jackson solvent bottles. Each standard was shaken eight times for one min each and used the next day. The three spiking levels were obtained by using 30 ml, 150 ml, and 750 ml of the water standard. All spikes were 750 ml in volume; the lower two levels required make-up water to achieve this volume.

Analysis of aqueous solutions prepared as PCB mixture or DDT isomer spikes. Separately prepared spiking solutions were analyzed for total available spike content as a methods check. The 3.00 liter

aqueous solution prepared as above was divided into 4 equal aliquots. Each aliquot was extracted three times with 75 ml cyclopentane each time. The solvent was dried with anhydrous Na_2SO_4 , concentrated, and analyzed by GLC.

Recovery of PCB or DDT from the membrane filter following spiking with aqueous solutions. Each spiked water sample was filtered through a membrane filter with water aspirator suction. Double-distilled water (100 ml) was used to wash down the walls of the funnel. The filter was extracted in a screw-capped 200-ml bottle with 3 portions of hexane (100, 10, and 10 ml) by mechanically shaking (30, 10, and 10 min each) in the presence of anhydrous Na_2SO_4 . The samples were concentrated and put through florisil columns utilizing the methylene chloride method (MILLS 1972) adapted to 1/10 size. GLC analyses followed.

Recovery of PCB or DDT from membranes spiked directly. The membranes to be dosed directly were wetted with water and rolled up in the mouth of a 200-ml extraction bottle. Standards of μl quantities (10-100) were dropped on the filters. Extraction with hexane, etc. was the same as that described above.

All spiking was done in triplicate and at three levels for each PCB mixture or DDT isomer tested. Levels were in multiples of five, the lowest giving full recorder scale detection at 100% recovery.

RESULTS AND CONCLUSIONS

Aroclor 1242 and Aroclor 1254 as well as p,p'-DDT were found to be efficiently adsorbed onto the surface of a cellulose triacetate membrane filter. shows results of the extraction of the eluted water spikes passing through the filter. The filter removed up to 60 µg of these species. The membrane shows promise for rapid removal of these chemical species from contaminated supplies not only for research or testing purposes but also for community water supplies. While a mechanical pump would be required for cleaning up large quantities of water, a simple gas sampling syringe with attached filter holder would suffice for testing purposes. Either a pure water sample would be taken or the purity could be checked in this manner in the field. The membrane filter could easily be stored and transported to the laboratory for analytical testing.

TABLE 1

Analysis of Membrane Effluent From a Water Spike

Compound or Mixture	Applied Quantity ¹	Adsorption
Aroclor 1242	40,000	98.0
Aroclor 1254	60,000	99.1
p,p'-DDE	2,000	97.9
<u>p</u> , <u>p</u> '-DDT	4,000	99.1

¹Dissolved in 750 ml water, flow rate of 200 ml/min.

On the other hand, desorption of these constituents from the membrane was found to be inefficient and inconsistent. Filter discs were first extracted with hexane in the presence of anhydrous Na₂SO₄. Low recoveries were obtained. The same discs were then extracted with acetonitrile in a tissue grinder. However, even with this dual treatment, the extraction ranged from 41 to 118% recovery (Table 2). Generally speaking, a higher recovery was obtained with the latter treatment. It is clear that additional work needs to be done to improve the consistency and efficiency of the extraction process if membrane filters are to be used for analytical purposes.

The water standards prepared as a spike were quantitatively analyzed. The Aroclor 1242 recovery averaged 79% for the 4 aliquots. Aroclor 1254 was recovered in two trials on an average of 82 and 79%. The p,p'-DDE and p,p'-DDT recoveries averaged 107 and 110%, respectively. Apparently, the Aroclors were adsorbed to the glass surface of the jug to a reasonably high degree (BELLAR et al. 1975). A subsequent hexane extraction of the jug for one trial for Aroclor 1254 gave a 7% recovery. These data were used to correct the quantity of PCB and DDT's present in the water spikes of the membrane filters.

These spikes were also applied directly to wetted membrane circles. The hexane extractions indicated a similar difficulty for recovering them (Table 3). Recoveries of individual samples of Aroclor 1242 ranged from 16 to 66% over the three levels of application, those of Aroclor 1254 from 32 to 92%. Those of Aroclor 1254 were higher at all spiking levels than those of Aroclor 1242. Data for the individual DDT isomers and aldrin are also given in Table 3. The

isomers p,p'-DDE and p,p'-DDT were studied at three levels while those of p,p'-TDE, o,p'-DDT, and aldrin were studied at only two. In all cases, recovery results were lower and even less consistent than those for the Aroclors.

TABLE 2

Recovery of Aroclors and DDT's from Membrane After Filtration of a Water Spike. Extraction of Membrane with Hexane and then Acetonitrile (1).

Extractn Solvent (2)	Applied Recovery Quant Trials ng 1 2 3	Ave %	Applied Quant ng	Recovery Trials 1 2 3	Ave %	
Compound	:Aroclor 1242		Aroclor 1254			
Hexane Aceton. Total	$\begin{array}{rrrrr} 400 & 43 & 13 & 31 \\ & 97 & 62 & 109 \\ \hline 140 & 75 & 140 \end{array}$	118	600	31 25 59 26 33 32 57 58 91	69	
Hexane Aceton. Total	2000 62 35 35 21 64 64 83 99 99	94	3000	26 3 46 39 81 37 65 84 83	77	
Hexane Aceton. Total	$\begin{array}{cccccccccccccccccccccccccccccccccccc$	78	15,000	26 6 18 18 36 21 44 41 39	41	
Compound	: <u>p</u> , <u>p</u> '-DDE		p,p'-DDT			
Hexane Aceton. Total	20 25 18 45 42 59 33 67 77 78	74	40	10 5 24 34 51 53 44 56 77	59	
Hexane Aceton. Total	$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	72	200	23 22 21 66 lost 59 89 - 80	85	
Hexane Aceton. Total	$\begin{array}{cccccccccccccccccccccccccccccccccccc$	54	1000	27 4 6 15 49 33 42 53 39	45	

⁽¹⁾ Data corrected for recovery of spikes from aqueous solutions.

⁽²⁾ Aceton. is acetonitrile.

TABLE 3
Recovery of Aroclors and DDT's from Membrane After Direct Spiking. Extraction of Membrane with Hexane.

Applied Quantity ng	Rec Tr:	cove ials 2	ry , % 3	Ave	Applie Quant: ng	ity	Tr	cove ials	<u>%</u>	Ave
Aroclor 1242					Aroclo	or 1	L254			
400 2000 10,000	16 31 38	66 44 48		42 40 36		300 000 000	53 65 92	32 58 56	52 63 75	46 62 71
p,p'-DDE					p,p'-1	DDT				
15 75 400	lost 13 26	48 9 30	80 45 24	64 22 27		40 200 000	lost 8 36	16 4 33	22 27 22	19 13 30
<u>p</u> , <u>p</u> '-TDE					<u>o,p</u> 'Dl	DT				
40 200	lost 8	26 3	35 29	31 13		40 200	lost 9	17 5	23 27	20 14
Aldrin										
5 25	lost 5	107 0	183 11	145 5						

Cellulose triacetate membrane filters exhibit a high degree of adsorption of organochlorine compounds. This quality provides potential uses in two areas, analysis and purification.

This work has shown that the use of cellulose triacetate membrane filters is an excellent adsorption method for the analysis of PCB's and DDT isomers in water. Using the method in the field would be simple with a minimum of equipment required. The used filters could easily and cheaply be stored and shipped to a laboratory for analysis. While desorption methods are not now efficient, further studies for their improvement are under investigation.

Purification of waters from PCB and organochlorine insecticides is of current ecological interest. SHTANNIKOV (1972) has studied flocculation, coagulation, and filter agents to this end. The use of aluminum sulfate and ferric sulfate as flocculating agents or

sand as a filtering medium were not successful even at higher pH's. The combination of aluminum sulfate, sand, and polyacrylamide did give water of high purity. Water plants gave limited results as a purification method. Ionites were more effective than activated carbon, while ion exchange polymers were fairly effective. SCHOLZ and ALTMANN (1971) studied the use of packed columns for removal of chlorinated hydrocarbons from drinking water. They reported removal of up to 73% applied DDT by activated carbon and polyethylene (both 2000 and 300,000 mol wt). Polyiosbutylene (10-15,000 mol wt) removed 88% of the applied DDT. On the other hand, this study has shown that cellulose triacetate membrane filters were very effective.

There are possibilities this type of filter could be applicable to air pollution problems.

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REFERENCES

- AHLING, B., and S. JENSEN: Anal. Chem. 42, 1483 (1970).
- BEDFORD, J. W.: Bull. Environ. Contam. and Toxicol. 12, 622 (1974).
- BELLAR, T. A., and J. J. LICHTENBERG: Water Quality Parameters, \underline{ASTM} \underline{STP} $\underline{573}$, 206 (1975).
- BERCK, B.: Anal. Chem. <u>25</u>, 1253 (1953).
- BIDLEMAN, T. F., and C. E. OLNEY: Bull. Environ. Contam. and Toxicol. 11, 442 (1974).
- BREIDENBACH, A. W., J. J. LICHTENBERG, C. F. HENKE, D. J. SMITH, J. W. EICHELBERGER, and H. STIERLI: U.S. Dept. of the Interior, FWPCA Publ. No. WP-22, Nov. (1966).
- CRAMER, J.: Atmos. Environ. $\underline{7}$, 241 (1973).
- ENVIRONMENTAL PROTECTION AGENCY: Methods for Organic Pesticides in Water and Wastewater, National Environ. Res. Center, Cincinnati, OH 45268 (1971).

- ENVIRONMENTAL PROTECTION AGENCY; Federal Register, Part IV, Dec. 24, 59566 (1975a).
- ENVIRONMENTAL PROTECTION AGENCY: Federal Register, Part IV, Dec. 24, 59587 (1975b).
- GESSER, H. D., A. CHOW, F. C. DAVIS, J. F. UTHE, and J. REINKE: Anal. Letters 4, 883 (1971).
- GIAM, C. S., H. S. CHAN, and G. S. NEFF: Anal. Chem. 47, 2319 (1975).
- JENSEN, S., A. G. JOHNELS, M. OLSSON, and G. OTTERLIND: Nature 224, 247 (1969).
- KAHN, L., and C. H. WAYMAN: Anal. Chem. 36, 1340 (1964)
- MILLS, P. A., B. A. BONG, LaV. R. KAMPS, and J. BURKE: JAOAC $\underline{55}$, 39 (1972).
- RISEBROUGH, R. W., P. RIECHE, D. B. PEAKALL, S. G. ḤERMAN, and M. N. KIRVEN: Nature 220, 1098 (1968).
- ROSEN, A. A., and E. M. MIDDLETON: Anal. Chem. <u>31</u>, 1729 (1959).
- SHTANNIKOV, E. V.: Gig. and Sanit. 9, 97-99 (1972). Water Res. Abstr. 7(21), 69 (Nov. 1, 1974) (5D-W74-11183).
- UTHE, J. F., J. REINKE, and H. GESSER: Environ. Letters $\underline{3}$, 117 (1972).
- UTHE, J. F., J. REINKE, and H. O'BRODOVICH: Environ. Letters 6(2), 103-115 (1974).
- WOODWELL, G. M., P. P. CRAIG, and H. A. JOHNSON: Science 174, 1101 (1971).